



Research Article

Development and Validation of Stability Stability-indicating HPLC Method for Related Substances Analysis of Fluorometholone in an Ophthalmic Solution

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Abstract

Anti-inflammatory ophthalmic solutions containing fluorometholone (FLM) combinations are commonly used, and very few stability-indicating liquid chromatographic methodshave been reported for their related substances. In this study, a simple HPLC method was developed and validated for the determination of fluorometholone impurities in an ophthalmic solution containing FLM and tetrahydrozoline hydrochloride. All impurities were separated by using gradient elution at a flow rate of 1.5 ml/min using a $\mu Bondapak$ C18 250 mm \times 4.6 mm, 5 μm (or equivalent) column kept at 40 °C with 20 μl injection volumes. The wavelength was 240 nm. This method validation included specificity, linearity, limit of detection (LOD), limit of quantification (LOQ), accuracy, precision, and robustness. The stability-indicating capability of this method was evaluated by performing forced degradation stress studies.

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Submitted: October 15, 2025 **Approved:** October 21, 2025 **Published:** October 22, 2025

How to cite this article: Sener C, Ergen H, Guleli M, Caliskan C. Development and Validation of Stability Stability-indicating HPLC Method for Related Substances Analysis of Fluorometholone in an Ophthalmic Solution. Arch Case Rep. 2025; 9(10): 326-332. Available from:

https://dx.doi.org/10.29328/journal.acr.1001168

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Keywords: Fluorometholone; RP-HPLC; Stress testing; Stability indicating; Related substances; Validation





Introduction

Fluorometholone (FLM), which is a corticosteroid that inhibits inflammation, and its combinations, such as FLM-ketorolac, FLM-tetrahydrozoline hydrochloride, are used in the case of allergic and inflammatory conditions in the eyes, eye redness/itching [1-3] (Figure 1).

The pharmacopeia survey finds that analytical methods are reported for solutions drops in British (BP) [4], US (USP)



Pharmacopeias, and cream, ophthalmic suspension, and ointment only in USP [5]. It is not reported related substances method is used for cream and ointment. Known impurities are not defined for ophthalmic solutions and suspensions in both USP and BP. For the FLM raw material monograph, only the USP identifies one of the known impurities, which is FLM-related compound A (Table 1). In this method, five FLM impurities (Table 1) were successfully separated from each other and detected. Analytical methods for the determination of fluorometholone in drugs were described, including HPTLC for impurity [6], UV spectrophotometry [7], TLC-spectrodensitometry [7], and HPLC for assay [2,9].

Experimental

Materials

Fluorometholone, impurities, and ophthalmic solution containing FLM and tetrahydrozoline hydrochloride sample were taken from commercial batches produced by the World Medicine Pharmaceutical Industry and Trade Inc. (Istanbul,



Table 1: Chemical structure of fluorometholone impurities.						
Name	Structure	IUPAC Name				
Deltamedrane (Imp A)	HO H H	$(6S, 8S, 10R, 11S, 13S, 14S, 17R) - 17 - acetyl - 11, 17 - dihydroxy - 6, 10, 13 - trimethyl - 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17 - dodecahydro - 3H - cyclopenta[\alpha]phenanthren - 3 - one$				
Bromidrine	HO H H H	$(6S,\!8S,\!9R,\!10S,\!11S,\!13S,\!14S,\!17R)\!-\!17\text{-acetyl-}9\text{-bromo-}11,\!17\text{-dihydroxy-}6,\!10,\!13\text{-trimethyl-}6,\!7,\!8,\!9,\!10,\!11,\!12,\!13,\!14,\!15,\!16,\!17\text{-dodecahydro-}3\textit{H-}\text{cyclopenta}[\alpha]\text{phenanthren-}3\text{-one}$				
FLM2	O H H H H H H H H H H H H H H H H H H H	$(6S, 8S, 10R, 13S, 14S, 17R) - 17 - acetyl - 17 - hydroxy - 6, 10, 13 - trimethyl-6, 7, 8, 10, 12, 13, 14, 15, 16, 17 - decahydro - 3H - cyclopenta [\alpha] phenanthren - 3 - one$				
FLM3	O H	(4aS,4bS,5aS,6aS,7R,9aS,11S)-7-acetyl-7-hydroxy-4a,6a,11-trimethyl-5a,6,6a,7,8,9,9a,9b,10,11-decahydrocyclopenta[1,2]phenanthro[4,4a-b]oxiren-2(4aH)-one				
1,2-Dihydrofluorometholone (Imp B)	HO HO	$(6S,9R,10S,11S,13S,14S,17R)-17-acetyl-9-fluoro-11,17-dihydroxy-6,10,13-trimethyl-1,2,6,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-3\emph{H}-cyclopenta[α] phenanthren-3-one$				

Turkey). Analytical reference standards used during development and validafollows and their purities given in parentheses; fluorometholone (99.32%) and impurities Deltamedrane (95.7%), 1,2-Dihydrofluorometholone (100.0%), Bromidrine (100.0%), FLM2 (96.8%), FLM3 (100.0%). Sodium hydroxide (NaOH), hydrochloric acid (HCl), hydrogen peroxide ($\rm H_2O_2$), triethylamine (Et $_3$ N), potassium dihydrogen phosphate, and o-phosphoric acid were purchased from Merck, methanol from J. T. Baker, and tetrabutylammonium hydroxide solution (40%) from Acros. HPLC-grade water (0.05 μ c) was produced by the Sartorius Stedim Biotech system.

Chromatographic conditions and preparation of solutions

Preparation of mobile phase A:

Mix 500 ml purified water and 500 ml methanol in a 1000 ml volumetric flask and adjust pH to 3.2 ± 0.05 with concentrated phosphoric acid and degase.

Preparation of mobile phase B:

Purified water: Methanol: Phosphoric acid (97: 3: 0.05) (v: v: v).

Related substances HPLC method: The HPLC method

developed for related substances analysis of fluorometholone was carried out on $\mu Bondapak$ (250 mm \times 4.6 mm, 5 μm) column with 20 μl injection volume at a wavelength of 240 nm on a Waters Alliance E2695 separation module equipped with a Waters 2489 photodiode array (PDA) detector and 2998 UV detector, an Empower-pro data handling system (Waters Corporation, Milford, MA, USA). Column and sample temperatures were 40 °C and 25 °C, respectively. The separation was employed using gradient elution based on the programsin Table 2. Methanol is used as a dilution solution, and all prepared solutions were filtered through a 0.45 μm PTFE filter.

Preparation of standard solution: Weigh accurately about 10.0 mg Fluorometholone into a 100 ml volumetric flask, add some dilution solution and sonicate in an ultrasonic bath until dissolved, complete to volume with dilution solution and mix (Stock solution 1). Transfer 5.0 ml of the obtained solution into a 50 ml volumetric flask, complete to volume with dilution solution, and mix well. Transfer 5.0 ml of the obtained solution into a 50 ml volumetric flask, complete to volume with dilution solution, and mix. Filter through a PTFE filter with a pore size of 0.45 μm and take an HPLC vial.

Preparation of system suitability solution: Weigh accurately 2.0 mg 1,2-Dihydroderivated (Imp B) into a 50 ml



Table 2: Program of the method.							
Time (min.)	Mobile Phase A (%)	Mobile Phase B (%)					
0	60	40					
20	80	20					
50	80	20					
55	90	10					
65	100	0					
75	100	0					
76	60	40					
85	60	40					

volumetric flask, add 30.0 ml stock solution 1, sonicate in an ultrasonic bath until dissolved, complete to volume with stock solution 1, and mix. Transfer 0.5 ml of the obtained solution into a 20 ml volumetric flask, complete to volume with stock solution 1, and mix well. Filter through a PTFE filter with a pore size of 0.45 μm and take an HPLC vial.

Preparation of the sample solution: Take $\sim\!\!1.0$ ml sample equivalent to 1.0 mg Fluorometholone into a 10 ml volumetric flask. Add some dilution solution and sonicate in an ultrasonic bath until dissolved, complete to volume with dilution solution, and mix. Filter through a PTFE filter with a pore size of 0.45 μm and take an HPLC vial.

Stress-testing and stability studies: The ophthalmic solution containing FLM and tetrahydrozoline hydrochloride sample were subjected to stress-testing under the following conditions: thermal degradation (standing at 60 °C 14 days), photolytic degradation under day-ligat 25 °C for 14 days, acidic hydrolysis (standing at 5.0 M HCl solution at 25 °C for 14 days and at 60 °C for 14 days), alkaline hydrolysis (standing at 5.0 M NaOH solution at 25 °C for 14 days and at 60 °C for 14 days), and oxidative degradation (standing at 30% $\rm H_2O_2$ solution at 25 °C for 14 days). The acidic, basic, and oxidative hydrolysis solutions were neutralized by using 5.0 M NaOH, 5.0 M HCl, and 30% tetrabutylammonium solutions. For each study, corresponding blank solutionanalyzed to determine the formed degradation impurities and.

Optimization of chromatographic conditions

To obtain a good resolution among impurities and FLM, different columns were tested having C18 stationary phases with particle sizes of $10.0~\mu m$ diameters of 4.6~mm and 3.9~mm, and column lengths of 250~mm and 300~mm. It was found that impurities and FLM were well retained and separated with μB ondapak C18 250 mm \times 4.6 mm, 5 μm column. Since the pH level of the mobile phase may affect the chromatographic behaviors, after several trials, an ideal pH was found 3.2 for good resolution. When the pH was not adjusted, the resolution was lost between Deltamedrane impurity and FLM peaks (Figure 2).

Results and discussion

Ation of HPLC-related substances method

The optimized HPLC method was validated according to the ICH Q2 (R2) guideline [9,10]. The validation parameters

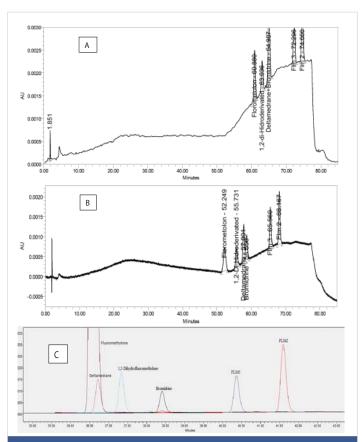


Figure 2: Optimization Chromatograms: A; column: ODS Hypersil 250 mm × 4.6 mm, 5 μm and mobile phase: Purified water: Methanol (1:1, v:v) adjusted pH:3.2, **B**; column: μBondapak C18 250 mm × 4.6 mm, 5 μm and mobile phase: Purified water: Methanol (1:1, v:v) adjusted pH:3.2, **C**; column: μBondapak C18 250 mm × 4.6 mm, 5 μm and mobile phase: Purified water: Methanol (1:1, v:v).

included system suitability, specificity, linearity, accuracy, precision (system, method, and intermediate precision), and robustness.

Specificity

In the specother peak was observed on the dilution and placebo solution chromatogram at the retimpurity peaks, which were all separated from each other and found spectrally pure (purity angle < purity threshold) (**Suppl. Mat. Table S1**). According to the results, the specificity of the developed method was found suitable for the determination of related impurities in FLM/ tetrahydrozoline hydrochloride ophthalmic solution.

System suitability

The system was approved to be suitable for use as the resolution between fluorometholone and 1,2-dihydrofluorometholone peaks was found to be 2.9 (\geq 2.0) in the chromatogram of the system suitability solution; symmetry factor was 0.9 (0.8 - 1.5), and theoretical plate count was 36869 (\geq 10000) obtained from the standard solution (Figure 3).

Linearity and range

The linearity of peak areas was checked using different



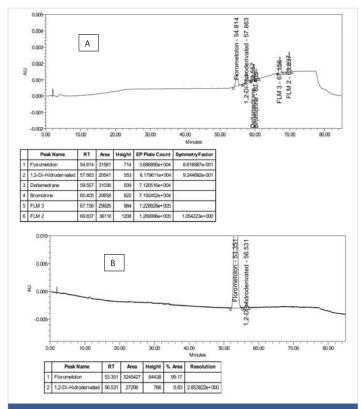


Figure 3: HPLC chromatograms: (A) mixture solution of fluorometholone and spiked with impurities at the limit concentration, (B) system suitability solution.

concentrations of standard solution from LOQ to 140%. The calibration curve has shown good linearity with the regression equation y = 31742569.104155x-154.473550 for FLM, and the correlation coefficient (r) was found to be 0.9998. The slope, intercept, and regression coefficient values of impurities were given in Table 3. The linearity graphics obtained from the Empower were given in the Suppl. Mat. S.3.2.3.

Limit of detection (LOD), limit of quantitation (LOQ), and relative response factor (RRF)

The LOD and LOQ were determined at a signal-to-noise ratio of 3:1 and 10:1, respectively. The determined LOD, LOQ, and RRF values for FLM and its impurities were reported in Table 4.

Accuracy

The accuracy was evaluated by measuring recovery through spiking known amounts of the impurities and FLM stinto to placebo containing tetrahydrozoline hydrochloride. Three different concentration levels 80%, 100%, and 120% were prepared and injected three times. Three samples were prepared at each level, and each sample was injected in triplicate. Good-to-excellent recoveries of impurities were achieved within the limit range of 80.0% – 120.0% levels (Table 5).

Precision and intermediate precision

Thaccount its repeatability and intermediate precision

Table 3: Results for linearity and range studies.								
Compound	Compound Concentration Corre (µg/ml) coeffic		Regression equation					
Fluorometholone	0.050 - 1.402	0.9998	y = 31742569.104155x - 154.473550					
Deltamedrane	0.053 - 1.487	0.9998	y = 29461192.055379x - 8.185844					
1,2-Dihydro- fluorometholone	0.052 - 1.463	0.9996	y = 19580893.497977x + 109.987692					
Bromidrine	0.051 - 1.414	0.9995	y = 20173296.309443x + 25.492796					
FLM 3	0.050 - 1.407	0.9999	y = 29216002.244713x + 50.449443					
FLM 2	0.052 - 1.450	0.9999	y = 35137462.922738x - 23.666053					

Table 4: Results for LOD, LOQ, and relative response factor.								
Compound	LO	D	LOQ		S/N		RRF	CRF
Compound	μg/ml	%	μg/ml	%	LOD	LOQ	KKF	CKF
Fluorometholone	0.015	0.015	0.050	0.050	3.70	10.64	1.0	1.0
Deltamedrane	0.016	0.016	0.053	0.052	3.04	10.55	0.6	1.6
1,2-Dihydro fluorometholone	0.016	0.016	0.052	0.053	3.01	10.05	0.9	1.1
Bromidrine	0.015	0.015	0.051	0.051	2.96	10.05	0.6	1.6
FLM 3	0.015	0.015	0.050	0.050	2.86	9.32	0.9	1.1
FLM 2	0.016	0.016	0.052	0.052	3.00	10.83	0.9	1.1

Table 5. Posulte for accuracy

Table 5: Results for accuracy.									
	Accuracy (n = 3)								
Compound	Spiked amount (%)	Conc. (µg/ml)	Conc. found (Mean, µg/ml)	RSD (%)	Recovery (%)				
	80	0.849	0.837	0.76	98.53				
Deltamedrane	100	1.062	1.056	0.26	99.39				
	120	1.274	1.275	0.66	100.01				
1,2-Dihydro-	80	0.836	0.839	0.60	100.34				
fluorometholone	100	1.045	1.032	1.44	98.77				
	120	1.254	1.247	0.92	99.47				
	80	0.808	0.811	1.63	100.37				
Bromidrine	100	1.010	0.987	0.82	97.72				
	120	1.212	1.211	0.96	99.95				
	80	0.804	0.808	0.65	100.56				
FLM 3	100	1.005	1.015	0.67	100.98				
	120	1.206	1.223	0.63	101.41				
	80	0.829	0.819	0.41	98.82				
FLM 2	100	1.036	1.035	0.49	99.91				
	120	1.243	1.237	0.62	99.49				

aspects. Repeatability was determined by injecting six individual preparations of a mixture solution of fluorometholone and spiked with impurities at the limit concentration solution in the same equipment on the same day, and the RSD of peak areas was found below 10.0%. To determine the precision mixture solution of Fluorometholone and its impurities at the limit concentration was prepared and injected six times. A sample solution was injected once to determine the amount of impurities. Six sample solutions spiked with impurities at of specification limit concentrations were prepared, and each was injected once. The intermediate precision was also checked by different analysts on different days using different equipment by working like precision. The RSD values of the contents of the detected impurities were calculated Tables 6,7).



Cable 6: Results for precision.								
Compound	System presision DCD (0/)		М	ethod Precision (n	= 6)			
Compound	System precision RSD (%)	Conc. (µg/ml)	Conc. found (µg/ml)	Recovery (%)	RSD (%)	Confidence interval at 95%		
Fluorometholone	0.81	_	-	-	-	-		
Deltamedrane	1.20	1.062	1.053	99.11	0.56	0.99 - 1.00		
1,2-Dihydrofluorometholone	1.56	1.045	1.048	100.24	0.56	1.00 - 1.01		
Bromidrine	1.91	1.045	1.032	98.75	0.78	0.98 - 1.00		
FLM 3	0.68	1.096	1.005	101.04	0.76	1.00 - 1.02		
FLM 2	0.82	1.036	1.031	99.50	0.19	0.99 - 1.00		
Maximum unknown impurity	-	-	-	-	2.63	0.16 - 0.17		
Total impurity	-	-	-	-	0.43	-		

Cable 7: Results for intermediate precision.								
Compound		Inte	rmediate Precision	(n = 6)		DCD (0/) (n = 12)		
Compound	Conc. (µg/ml)	Conc. found (µg/ml)	Recovery (%)	RSD (%)	Confidence interval at 95%	RSD (%) (n = 12)		
Fluorometholone	_	_	-	-	_	-		
Deltamedrane	1.034	1.009	100.41	0.51	1.00 - 1.01	0.85		
1,2-Dihydrofluorometholone	1.030	1.039	100.85	0.62	1.00 - 1.02	0.65		
Bromidrine	1.050	1.045	99.34	1.01	0.98 - 1.00	0.91		
FLM 3	1.050	1.051	100.08	0.51	1.00 - 1.01	0.79		
FLM 2	1.045	1.045	100.00	0.27	1.00 – 1.00	0.34		
Maximum unknown impurity	-	-	-	3.03	0.15 - 0.16	3.22		
Total impurity	-	_	_	0.24	-	0.35		

able 8: Results for robustness.								
		Column te	mperature	pH of mobile phase A				
Compound	% Variation for Using Different Column	% Variation for 38 °C	% Variation for 42°C	% Variation for pH 3.15	% Variation for pH 3.25			
Fluorometholone	1.35	0.06	0.80	0.59	0.35			
1,2-Dihydrofluorometholone	0.39	0.07	1.54	0.12	0.39			
Deltamedrane	2.38		0.77	0.53	1.88			
Bromidrine 0.26		1.38	0.60	0.12	0.80			
FLM 3	0.69	0.16	1.45	1.13	2.26			
FLM 2	3.57	1.19	0.84	0.13	2.57			

	S	tandard solution at 5	°C	Si	tandard solution at 25	5°C
Compound	% Variation for 6 hours	% Variation for 24 hours	% Variation for 48 hours	% Variation for 6 hours	% Variation for 24 hours	% Variation for 48 hours
Fluorometholone	2.06	2.72	1.69	0.76	0.14	1.69
1,2-Dihydrofluorometholone	0.00	2.63	1.34	0.15	0.43	1.01
Deltamedrane	0.81	2.51	2.87	0.10	1.37	2.04
Bromidrine	0.49	0.63	0.54	0.30	1.10	0.78
FLM 3	0.73	0.68	1.36	1.62	1.50	0.60
FLM 2	0.41	2.48	2.29	0.85	2.25	2.57
Maximum unknown impurity	-	-	_	-	_	_
Total impurity	_	_	_	-	_	_

Robustness

Into investigate the robustness of the method, minor but important changes in method parameters (column temperature by ±2 °C, pH of mobile phase A by ±0.05 units, using a different column) were made, and standard solutions spiked with impurities at the specification limit were tested. % Variation was calculated, and no significant difference was found between initial and altered conditions (Table 8).

Stabilityyof standard and sample solutions

Solution stability was also evaluated by monitoring the peak area response. Impurity spiked standard and sample solutions were analyzed right aftertheir preparation, 6, 24, and 48 hours after at 5 °C and at room temperature. Standard and sample solutions were stable for 48 hours since the variations were below 10.0% Tables 9,10).

Stress testing

 $Stress\,testing\,helps\,to\,determine\,the\,stability\,of\,the\,molecule$ that is exposed to degradation under different conditions and to validate the stability-indicating power of the analytical methods used. In this study, the degradation profile of FLM/ tetrahydrozoline hydrochloride ophthalmic solution was monitored by applying stress-testing conditions mentioned above in Section 2.3. According to the results of stress-testing studies (Table 11), under thermal, acidic, alkaline, oxidative,



Table 10:	Results for	sample so	lution stability.

	Sample solution at 5 °C			Sample solution at 25 °C			
Compound	% Variation for 6 hours	% Variation for 24 hours	% Variation for 48 hours	% Variation for 6 hours	% Variation for 24 hours	% Variation for 48 hours	
Fluorometholone	0.91	0.93	2.04	0.14	3.37	4.48	
1,2-Dihydrofluorometholone	4.79	1.08	0.80	0.85	1.74	1.55	
Deltamedrane	1.83	1.42	1.89	2.28	0.72	2.38	
Bromidrine	7.03	1.73	0.11	2.12	2.80	0.79	
FLM 3	0.06	1.00	1.51	1.20	4.64	0.52	
FLM 2	2.24	0.18	1.40	0.44	2.18	1.46	
Maximum unknown impurity	0.51	0.70	9.16	0.40	4.08	4.50	
Total impurity	2.96	0.12	0.77	0.55	2.08	0.34	

Tab	le 11	L: Resul	ts of str	ess-testii	ng studies.
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Compound	Untreated sample	Hydrolysis						
		Acidic		Basic		Oxidative degradation	Thermal degradation	Photolytic degradation
		25 °C, 14 th day	60 °C, 14 th day	25 °C, 14 th day	60 °C, 14 th day	uegrauation	uegrauation	uegrauation
Deltamedrane	ND	ND	ND	ND	ND	ND	ND	ND
1,2-Dihydrofluoro- metholone	ND	ND	ND	ND	ND	ND	ND	ND
Bromidrine	ND	ND	ND	ND	0.12	ND	ND	ND
FLM 3	ND	ND	ND	ND	ND	ND	ND	ND
FLM 2	ND	ND	ND	ND	ND	ND	ND	ND
Maximum unknown impurity	0.13	0.17	0.57	0.16	0.74	36.79	0.15	0.14
Total impurity (%)	0.24	0.28	2.14	0.29	2.53	37.84	0.31	0.25
Assay (%)	100.54	99.76	99.02	100.48	97.82	61.35	100.76	100.39

and photolytic conditions, known impurities have not been observed except for alkaline hydrolysis, standing at 60 °C for 14 days. The most degradation occurred in oxidative degradation, and it was noted that 36.79% unknown impurity formed. It was seen that there was an increase in the unknown impurities in all degradations.

Conclusion

Stabilitystability-indicating HPLC-related method was developed, validated, and used during analyses of stability samples of Fluorometholone/ tetrahydrozoline hydrochloride ophthalmic solution. The method was validated according to International Conference on Harmonization (ICH) guideline and considered simple, sensitive, selective, linear, precise, accurate, robust, andstable for the determination of FLM impurities in its pharmaceutical formulation. The proposed methods were validated and could be used for routine analysis in quality control laboratories. Many parameters, such as choosing column, adjusting pH, played a critical rolein the retention time of the related compounds and their resolution from each other. The sample has reached maximum degeneration in oxidation degradation and was found sensitive to acidic and basic hydrolysis at 60 °C. Under other degradation conditions, there was very little degradation. In fact, there was almost no decay under the light degradation. In the literature, it was seen that there was no reported stability-indicating method for the determination of fluorometholone impurities. So, this method could be a guide for the determination of impurities of all formulations containing fluorometholone.

Acknowledgment

This work was supported by the World Medicine Pharmaceutical Industry and Trade Inc. The authors are thankful to the company for providing the necessary instrumental facilities and chemicals to carry out the research work.

Supporting information: Supporting information accompanies this paper.

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